

Complex simulations

Most tutorials are about ubiquitine (also today's)



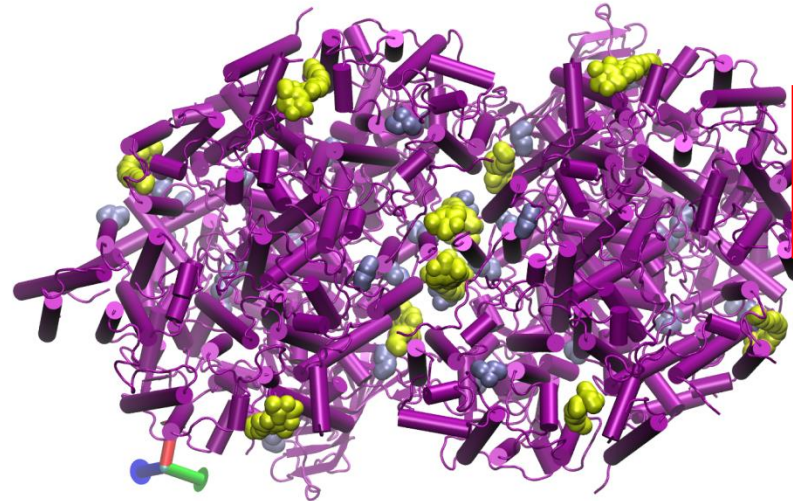
- Single protein chain
- No cofactors
- Easy to set up

What if your system looks more like this?



Preparing your structure for simulating

- Dimer!



- Add missing residues
- Add missing heteroatoms
- Delete all detergent molecules

```
REMARK 465  
REMARK 465 MISSING RESIDUES  
REMARK 465 THE FOLLOWING RESIDUES WERE NOT LOCATED IN THE  
REMARK 465 EXPERIMENT. (M=MODEL NUMBER; RES=RESIDUE NAME; C=CHAIN  
REMARK 465 IDENTIFIER; SSSEQ=SEQUENCE NUMBER; I=INSERTION CODE.)  
REMARK 465
```

```
REMARK 465 MET A 1  
REMARK 465 THR A 2  
REMARK 465 THR A 3  
REMARK 465 THR A 4  
REMARK 465 LEU A 5  
REMARK 465 GLN A 6  
REMARK 465 ARG A 7  
REMARK 465 ARG A 8  
REMARK 465 GLU A 9  
REMARK 465 SER A 10  
REMARK 465 ASN C 19  
REMARK 465 SER C 20  
REMARK 465 ILE C 21
```

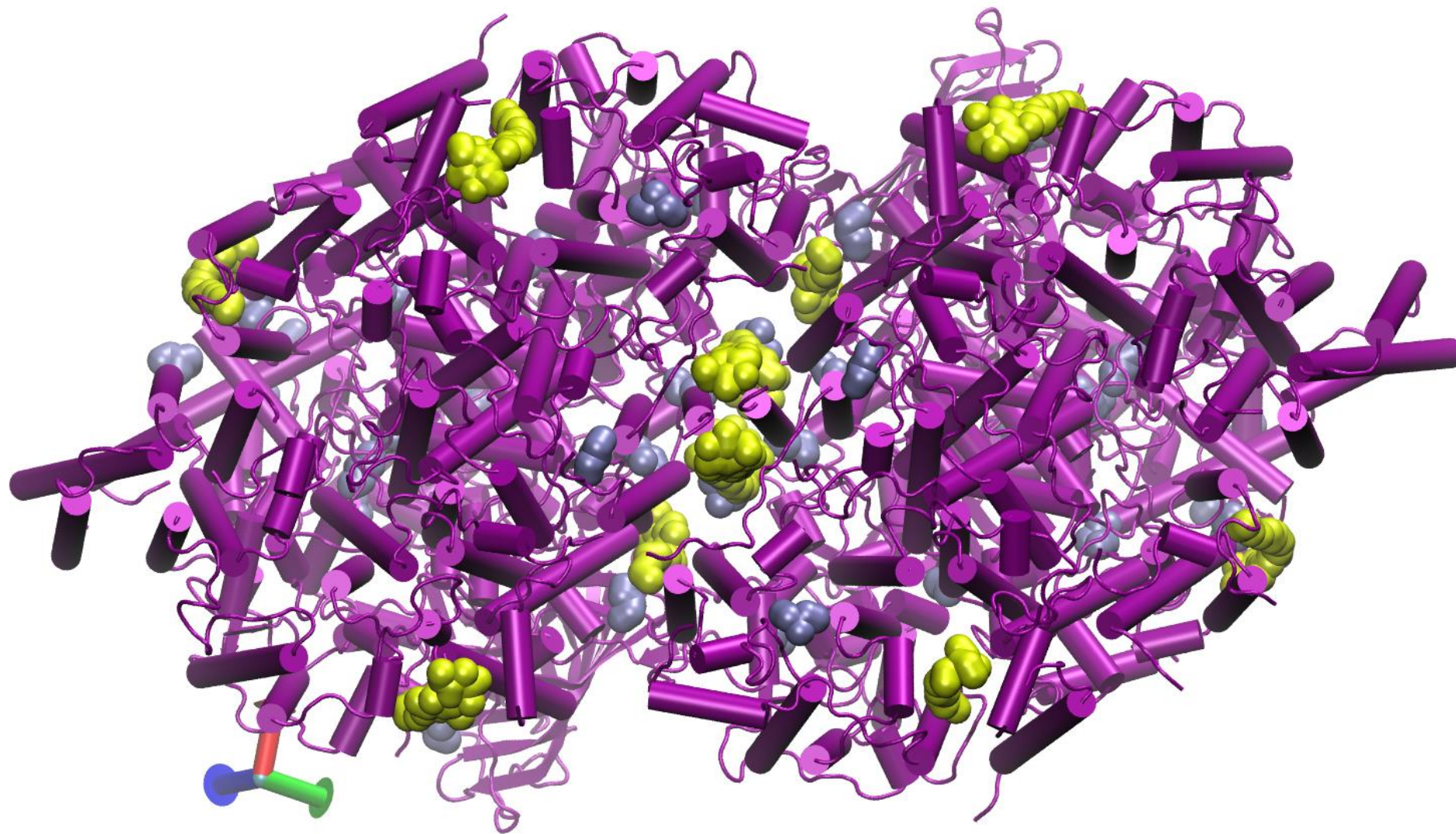
3arc.pdb

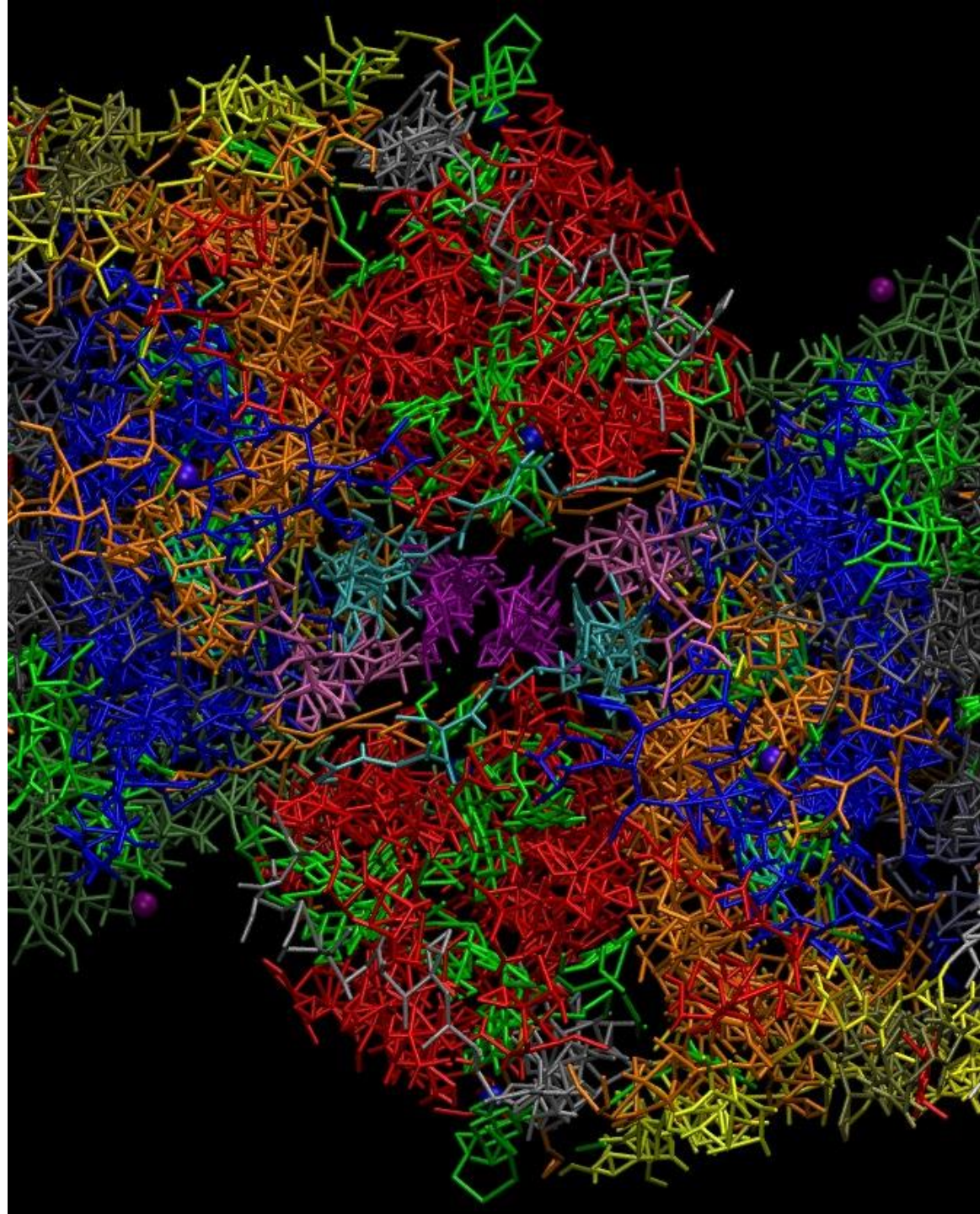
```
REMARK 610  
REMARK 610 MISSING HETEROATOM  
REMARK 610 THE FOLLOWING RESIDUES HAVE MISSING ATOMS (M=MODEL NUMBER;  
REMARK 610 RES=RESIDUE NAME; C=CHAIN IDENTIFIER; SSSEQ=SEQUENCE NUMBER;  
REMARK 610 I=INSERTION CODE):
```

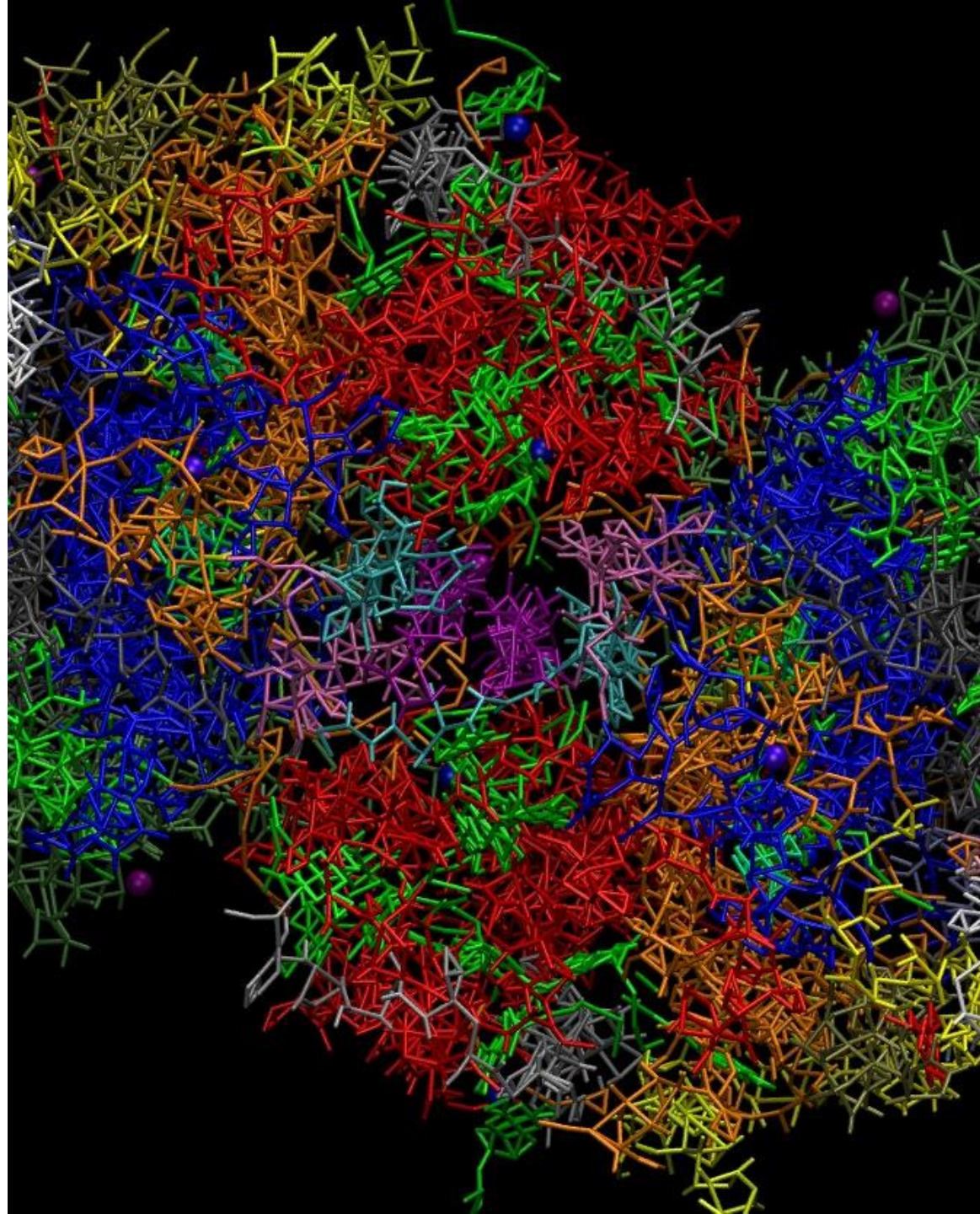
```
REMARK 610 M RES C SSSEQI
```

```
REMARK 610 LMG A 751  
REMARK 610 LMG B 669  
REMARK 610 DGD C 657  
REMARK 610 DGD C 660  
REMARK 610 DGD C 661  
REMARK 610 LMG C 729  
REMARK 610 LMG C 776  
REMARK 610 DGD D 755  
REMARK 610 SQD D 768  
REMARK 610 LMG D 692  
REMARK 610 HTG D 726  
REMARK 610 LHG E 772  
REMARK 610 DGD H 663  
REMARK 610 LMG Z 784  
REMARK 610 LMG a 751  
REMARK 610 LMG b 669  
REMARK 610 LMT b 791  
REMARK 610 DGD c 657  
REMARK 610 DGD c 660  
REMARK 610 DGD c 661  
REMARK 610 LMG c 729
```

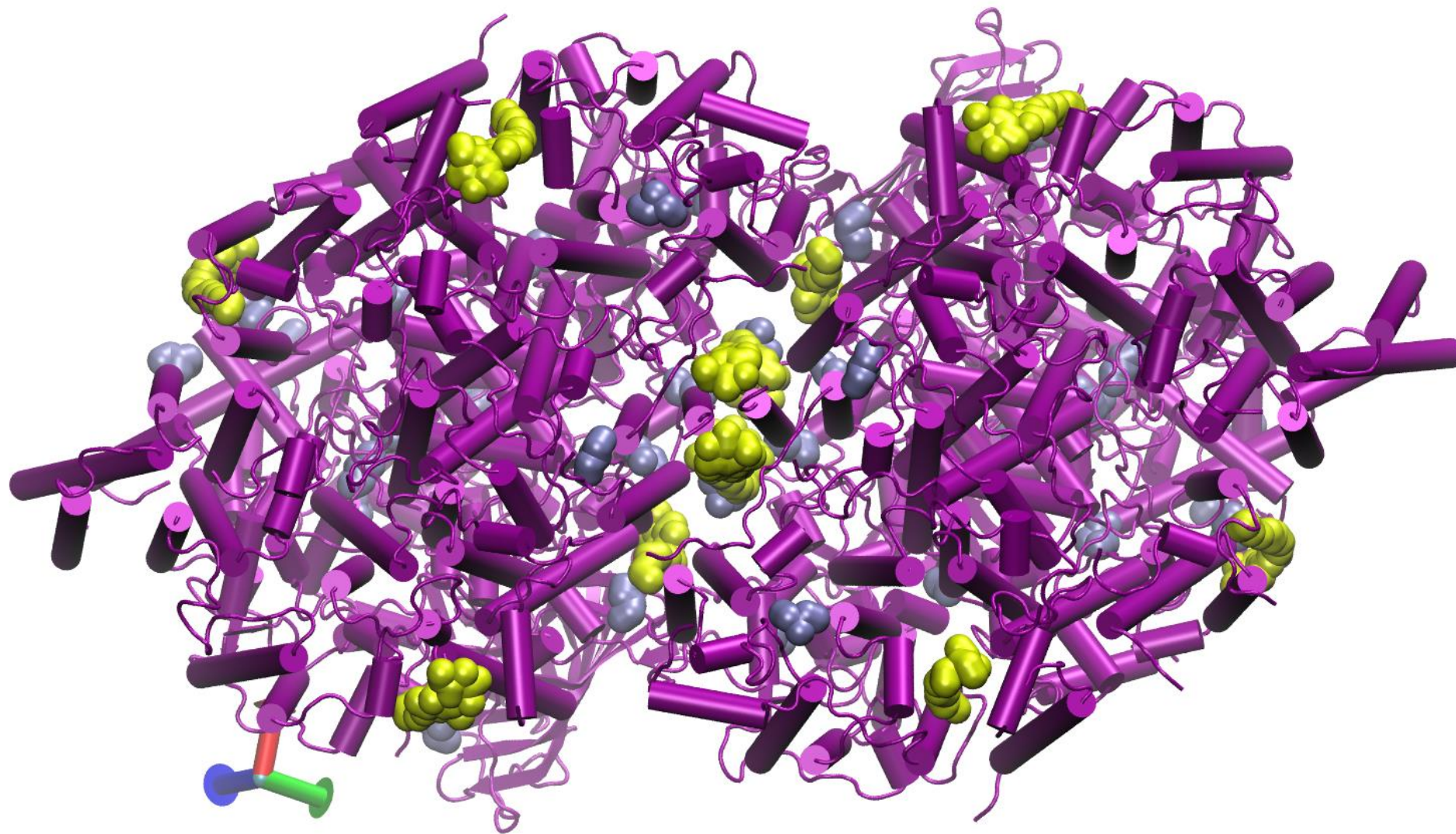
Detergents in crystal structure



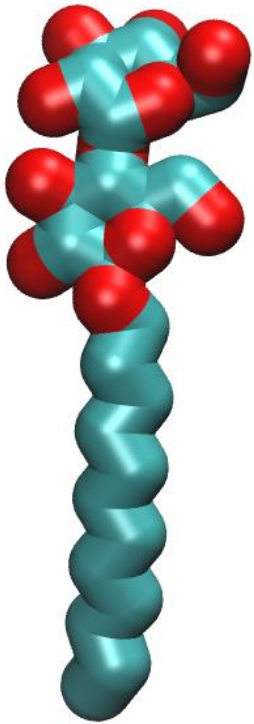




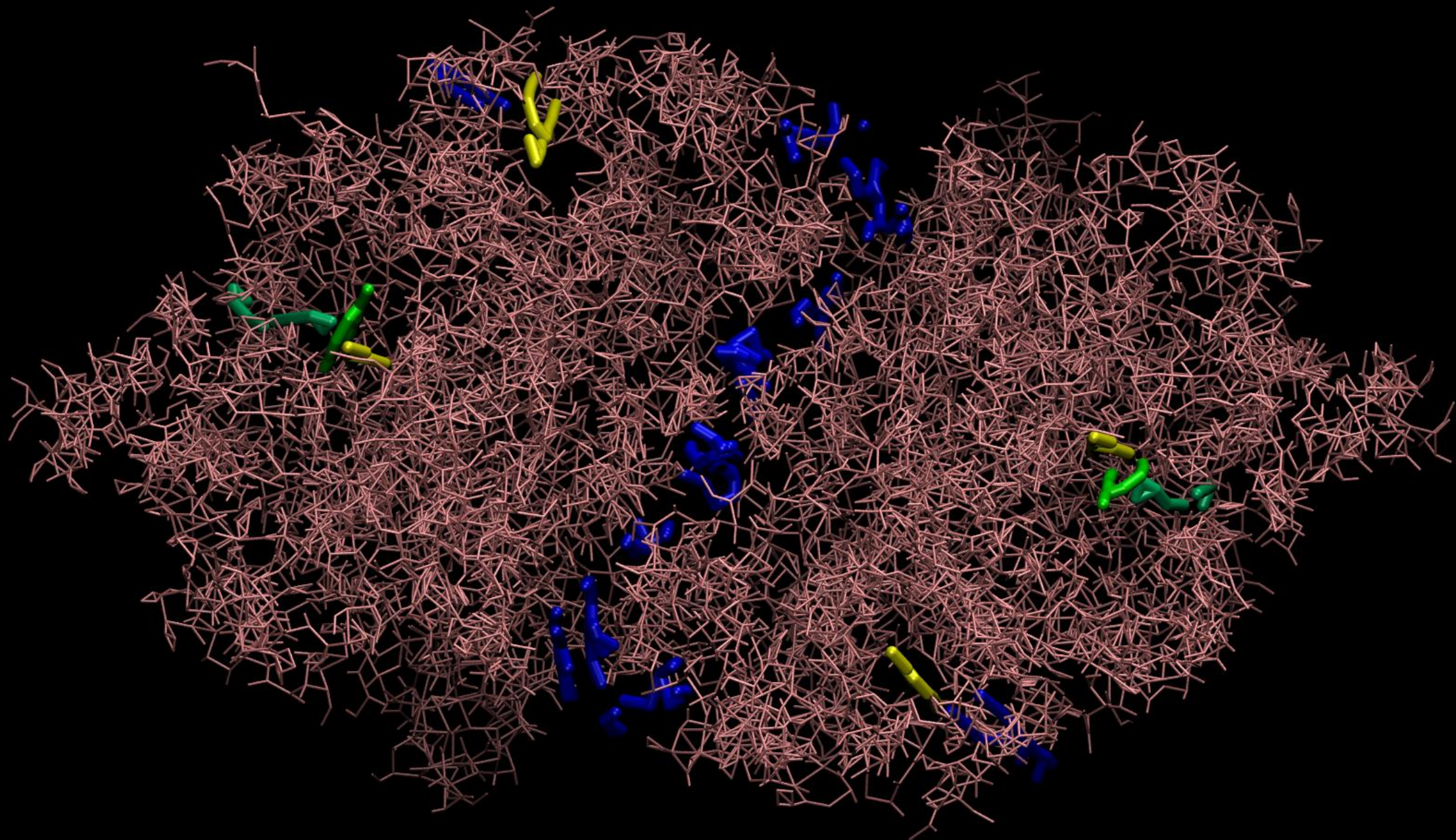
Detergents in crystal structure



Suprise suprise



DODECYL-BETA-D-
MALTOSIDE →
looks like a lipid



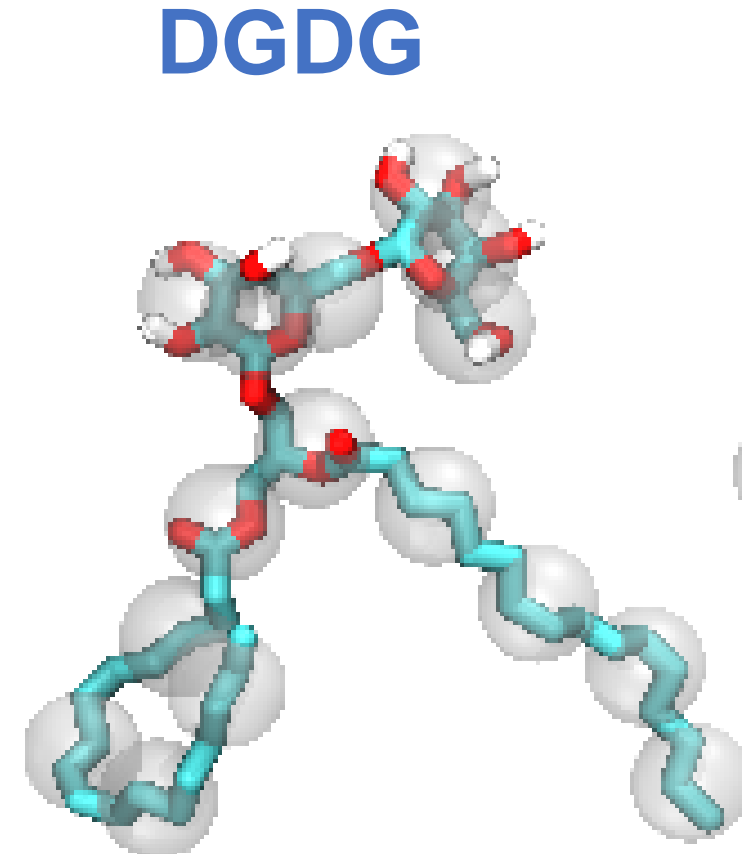
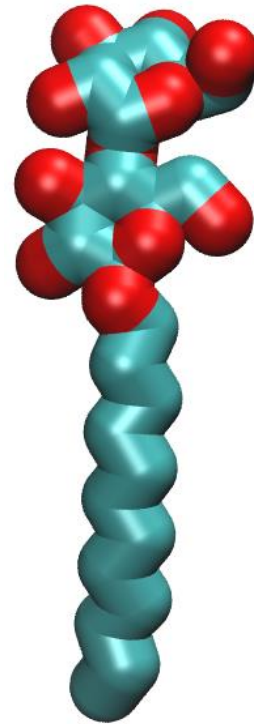
Where should I replace my detergents with?

Lipid that is the most similar?

Look in older structures.

Look in literature

→ No real good answer



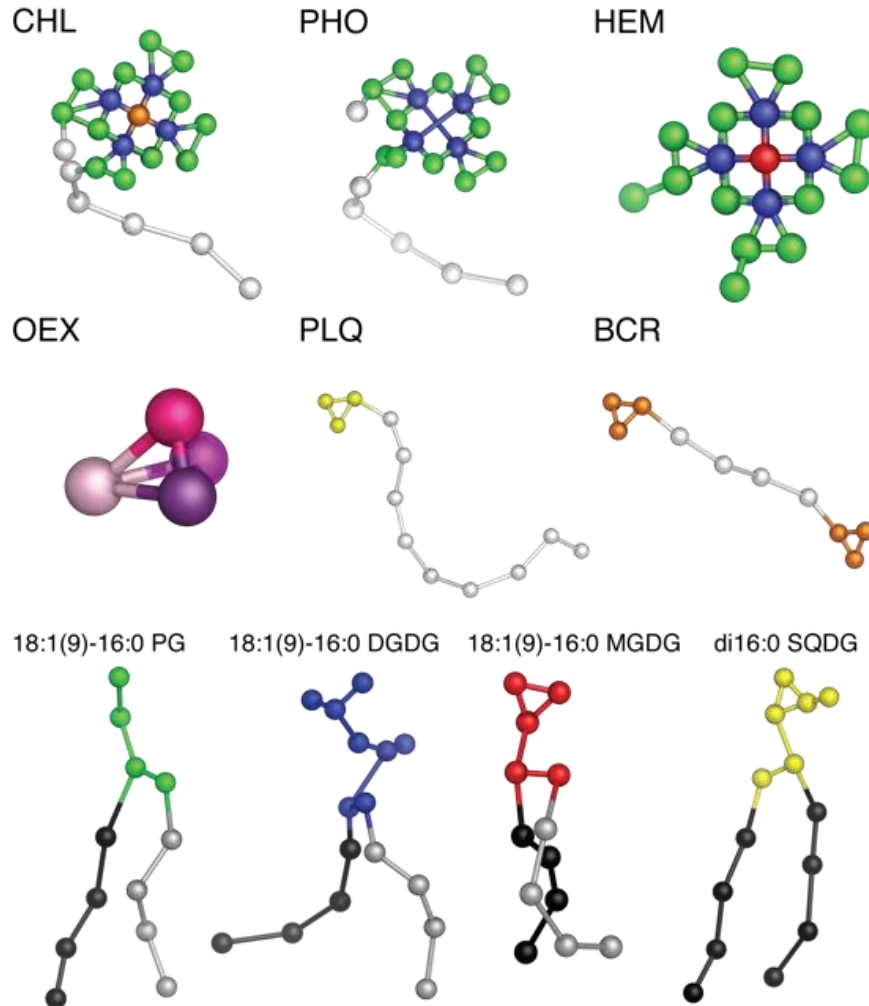
Coarse graining

- Split PDB
 - Protein → Martinize.py
 - Be careful Backward.py can also CG your protein, but it does not make an Elnedyn network, but most importantly the mapping of the backbone bead is different for the Elnedyn version
 - Cofactor → Backward.py

Embed in a bilayer

- Insane
 - OPM database → contains calculated position of the bilayer

Parameterization



PSII dimer (cyanobacteria)#

2x19 protein subunits

cofactors per monomer:

chlorophyll A (35)

pheophytin (2)

heme (2)

β -carotene (12)

plastoquinone (2)

ox. evolving compl. (1)

lipid composition:

18:1-16:0 PG (10%)

18:1-16:0 DGDG (25%)

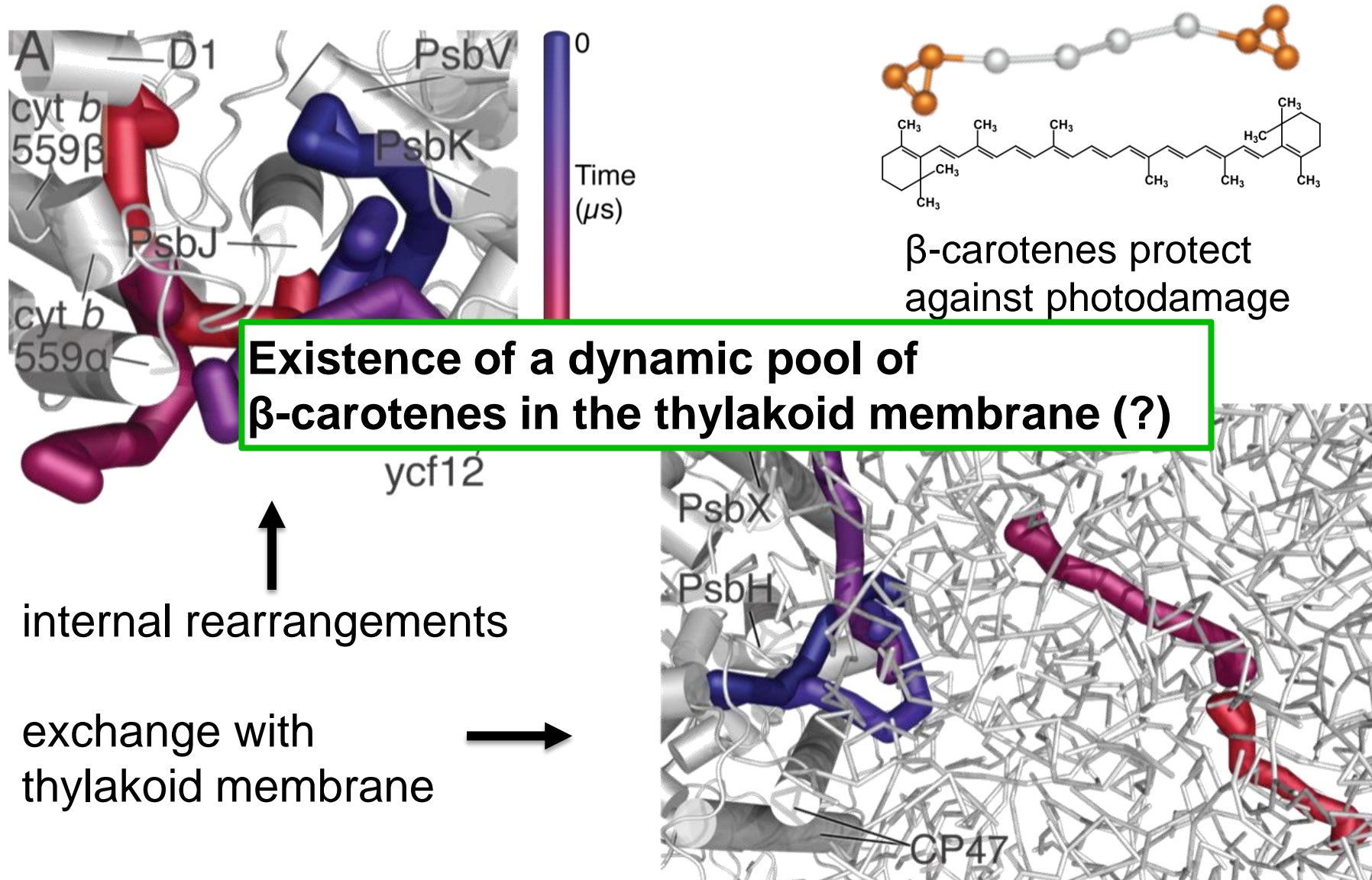
18:1-16:0 MGDG (40%)

18:1-16:0 SQDG (15%)

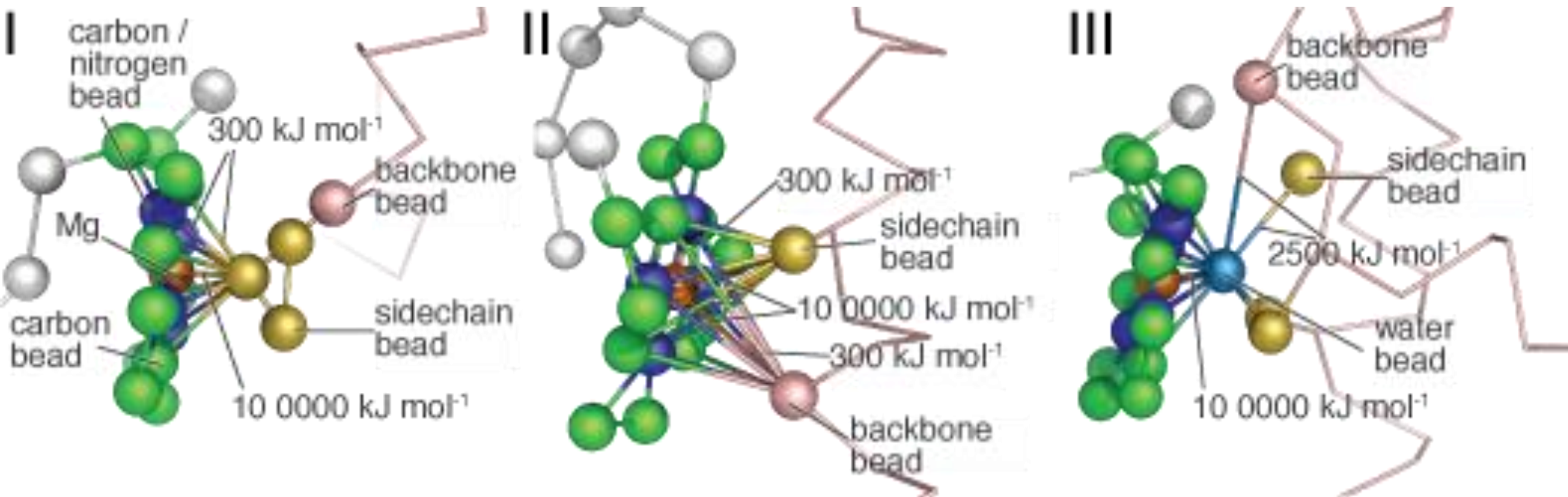
di16:0 SQDG (10%)

+ co-crystall. lipids (> 40 total)

β -carotenes are very mobile

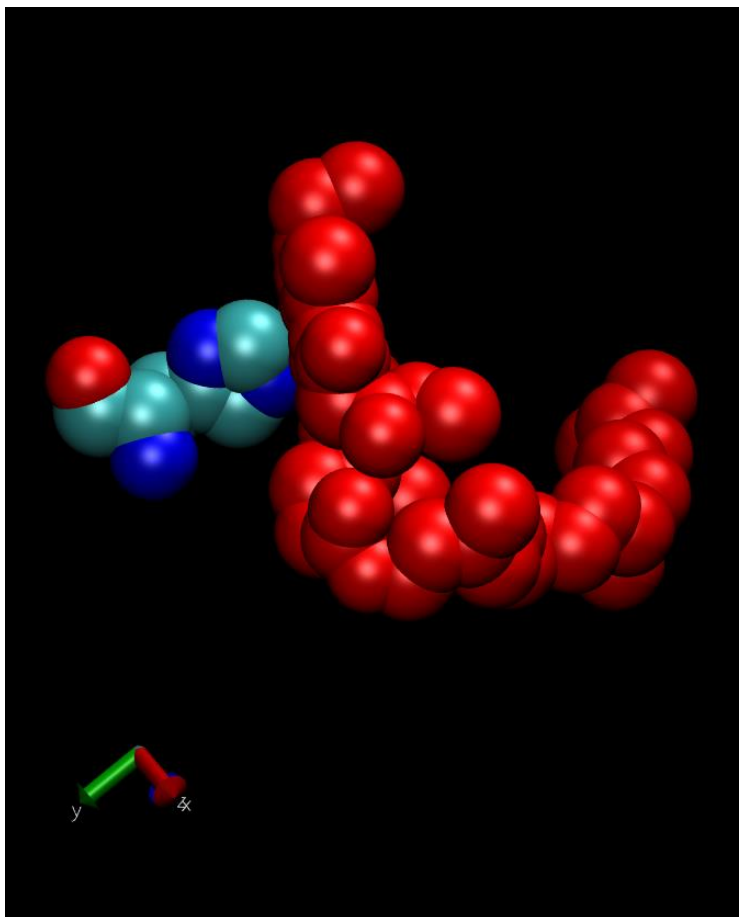


Chlorophylls are bound in 3 different ways

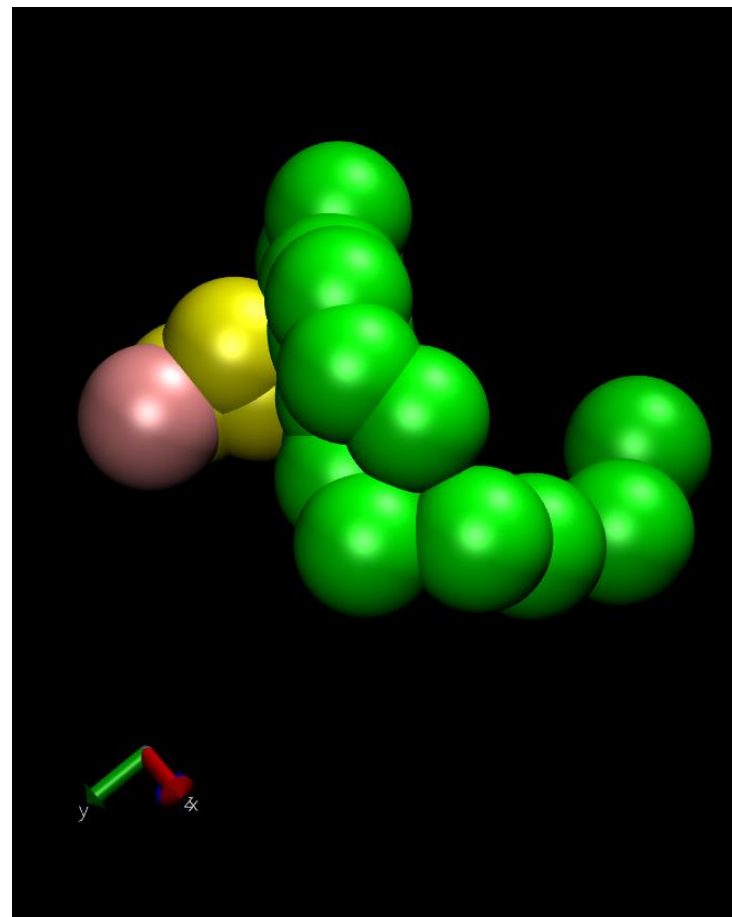


CG a bit too fat...

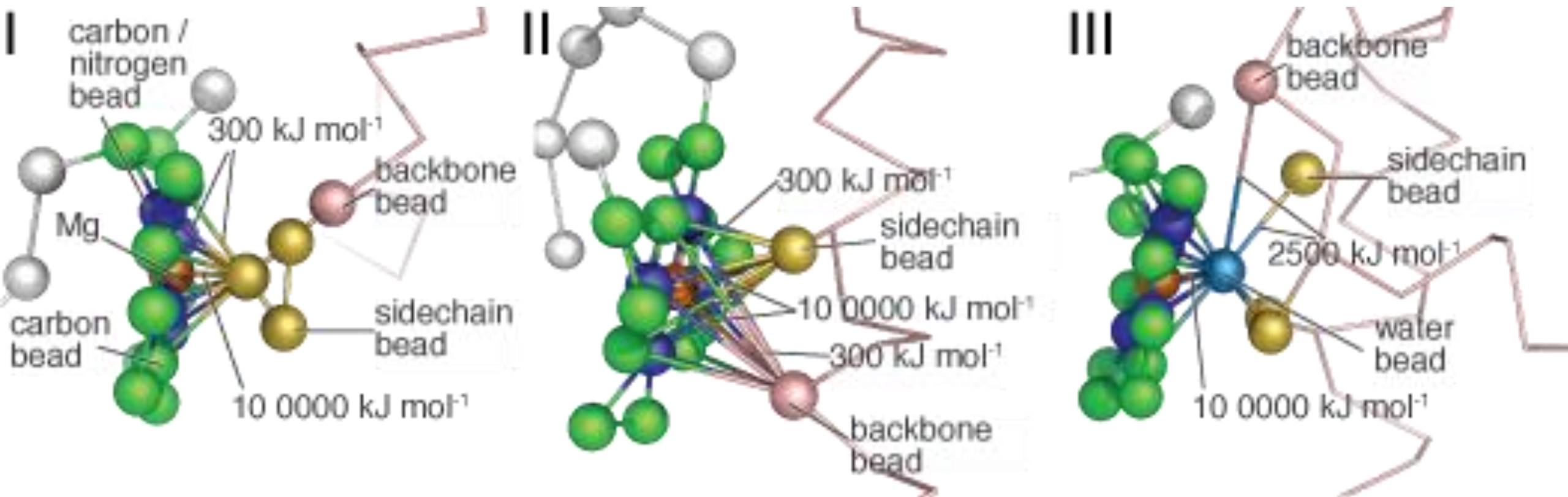
Fine grained



Coarse grained



Chlorophylls are bound in 3 different ways



Martinize.py → a itp file for every chain and cofactor

```
./Protein_T.itp
./Protein_Y.itp
./Protein_I.itp
./Protein_A.itp
./Protein_M.itp
./Protein_C.itp
./Protein_J.itp
./Protein_F.itp
./Protein_D.itp
./Protein_U.itp
./Protein_E.itp
./Protein_O.itp
./Protein_K.itp
./Protein_H.itp
./Protein_X.itp
./Protein_V.itp
./Protein_Z.itp
./Protein_L.itp
./Protein_B.itp
11:13:18@~/parameter/PS:
```

```
Chlorophyll-A Coarse-grained
; Modified CLA_CG topology JANUARY 2014, modified dir; De Jong et al ...
; MARTINI (elndyn22) Coarse Grained topology file for "Prote
; Created by py version 2.2
; Using the following options: -f monomer_no_COfactor.pdb -c
; Sequence:
; MTTTLQRRESANLWERFCNWTSTDNRLYVGWFGVIMIPTLLAATICFVIAFIAAPPVC
; GSFSDGMPLGISGTFNFMIVFQAEHNILMHPFHQLGVAGVFGGALFCAMHGSLVSSLIF
; HERNAHNFPDLA
; Secondary Structure:
; CCCCCSSCC1111H2222TCTTSSCC1111HHHHHHHHHHHHHHH2222CCCC
; TCCCCSCCEE1111HHHHHHH2222CCCC1111HHHHHHHHHHHHHHH2222TCCC
; TTTTTCSSCCCC
[ moleculetype ]
; Name Exclusions
Protein_A 1
[ atoms ]
1 Qd 1 MET BB 1 1.0000 ; C
2 C5 1 MET SC1 2 0.0000 ; C
3 P5 2 THR BB 3 0.0000 ; C
4 P1 2 THR SC1 4 0.0000 ; C
5 P5 3 THR BB 5 0.0000 ; C
6 P1 3 THR SC1 6 0.0000 ; C
7 P5 4 THR BB 7 0.0000 ; C
8 P1 4 THR SC1 8 0.0000 ; C
9 P5 5 LEU BB 9 0.0000 ; S
10 C1 5 LEU SC1 10 0.0000 ; S
11 P5 6 GLN BB 11 0.0000 ; C
12 P4 6 GLN SC1 12 0.0000 ; C
13 P5 7 ARG BB 13 0.0000 ; C
14 N0 7 ARG SC1 14 0.0000 ; C
15 Qd 7 ARG SC2 15 1.0000 ; C
16 P5 8 ARG BB 16 0.0000 ; C
17 N0 8 ARG SC1 17 0.0000 ; C
18 Qd 8 ARG SC2 18 1.0000 ; C
19 P5 9 GLU BB 19 0.0000 ; S
20 Qa 9 GLU SC1 20 -1.0000 ; S
21 P5 10 SER BB 21 0.0000 ; S
22 P1 10 SER SC1 22 0.0000 ; S
23 P4 11 ALA BB 23 0.0000 ; C
24 P5 12 ASN BB 24 0.0000 ; C
25 P5 12 ASN SC1 25 0.0000 ; C
26 Nd 13 LEU BB 26 0.0000 ; 1
27 C1 13 LEU SC1 27 0.0000 ; 1
28 Nd 14 TRP BB 28 0.0000 ; 1
29 SC4 14 TRP SC1 29 0.0000 ; 1
30 SNd 14 TRP SC2 30 0.0000 ; 1
31 SC5 14 TRP SC3 31 0.0000 ; 1
32 SC5 14 TRP SC4 32 0.0000 ; 1
33 Na 15 GLU BB 33 0.0000 ; 1
```

```
Chlorophyll-A Coarse-grained
; Modified CLA_CG topology JANUARY 2014, modified dir; De Jong et al ...
; MARTINI (elndyn22) Coarse Grained topology file for "Prote
; Created by py version 2.2
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; Sequence:
; MTTTLQRRESANLWERFCNWTSTDNRLYVGWFGVIMIPTLLAATICFVIAFIAAPPVC
; GSFSDGMPLGISGTFNFMIVFQAEHNILMHPFHQLGVAGVFGGALFCAMHGSLVSSLIF
; HERNAHNFPDLA
; Secondary Structure:
; CCCCCSSCC1111H2222TCTTSSCC1111HHHHHHHHHHHHHHH2222CCCC
; TCCCCSCCEE1111HHHHHHH2222CCCC1111HHHHHHHHHHHHHHH2222TCCC
; TTTTTCSSCCCC
[ moleculetype ]
; Name Exclusions
Protein_A 1
[ atoms ]
1 SP3 1 CLA NA 1 -0.25
2 SC3 1 CLA CHA 2 0.00
3 SP3 1 CLA NB 3 -0.25
4 SC3 1 CLA CHB 4 0.00
5 SP3 1 CLA NC 5 -0.25
6 SC3 1 CLA CHC 6 0.00
7 SP3 1 CLA ND 7 -0.25
8 SC3 1 CLA CHD 8 0.00
9 SC3 1 CLA C1A 9 0.00
10 SC3 1 CLA C2A 10 0.00
11 SC3 1 CLA C1B 11 0.00
12 SC3 1 CLA C2B 12 0.00
13 SC4 1 CLA C1C 13 0.00
14 SNa 1 CLA C2C 14 0.00
15 Na 1 CLA T1 15 0.00
16 SC3 1 CLA C1D 16 0.00
17 SC3 1 CLA C2D 17 0.00
18 Na 1 CLA T2 18 0.00
19 C3 1 CLA T3 19 0.00
20 C1 1 CLA T4 20 0.00
21 C1 1 CLA T5 21 0.00
22 C1 1 CLA T6 22 0.00
23 SQ0 1 CLA MG 23 1.00
[ bonds ]
; bonds between beads
; The bonds are finetuned against a backmapped gromos;
1 9 1 0.273 30000.0
9 10 1 0.298 23000.0
1 10 1 0.318 25000.0
3 11 1 0.274 25000.0
3 12 1 0.319 23000.0
3 12 1 0.29 21000.0
5 13 1 0.24 25000.0
5 14 1 0.355 21000.0
6 14 1 0.28 25000.0
13 14 1 0.29 20000.0
14 15 1 0.305 10000.0
```

```
If using this topology, please site:
; Modified CLA_CG topology JANUARY 2014, modified dir; De Jong et al ...
; MARTINI (elndyn22) Coarse Grained topology file for "Prote
; Created by py version 2.2
; Using the following options: -f monomer_no_COfactor.pdb -c
; Sequence:
; MTTTLQRRESANLWERFCNWTSTDNRLYVGWFGVIMIPTLLAATICFVIAFIAAPPVC
; GSFSDGMPLGISGTFNFMIVFQAEHNILMHPFHQLGVAGVFGGALFCAMHGSLVSSLIF
; HERNAHNFPDLA
; Secondary Structure:
; CCCCCSSCC1111H2222TCTTSSCC1111HHHHHHHHHHHHHHH2222CCCC
; TCCCCSCCEE1111HHHHHHH2222CCCC1111HHHHHHHHHHHHHHH2222TCCC
; TTTTTCSSCCCC
[ moleculetype ]
; Name Exclusions
Protein_A 1
[ atoms ]
1 SP3 1 PHO NA 1 0.00
2 SC3 1 PHO CHA 2 0.00
3 SP3 1 PHO NB 3 0.00
4 SC3 1 PHO CHB 4 0.00
5 SP3 1 PHO NC 5 0.00
6 SC3 1 PHO CHC 6 0.00
7 SP3 1 PHO ND 7 0.00
8 SC3 1 PHO CHD 8 0.00
9 SC3 1 PHO C1A 9 0.00
10 SC3 1 PHO C2A 10 0.00
11 SC3 1 PHO C1B 11 0.00
12 SC3 1 PHO C2B 12 0.00
13 SC4 1 PHO C1C 13 0.00
14 SNa 1 PHO C2C 14 0.00
15 Na 1 PHO T1 15 0.00
16 SC3 1 PHO C1D 16 0.00
17 SC3 1 PHO C2D 17 0.00
18 Na 1 PHO T2 18 0.00
19 C3 1 PHO T3 19 0.00
20 C1 1 PHO T4 20 0.00
21 C1 1 PHO T5 21 0.00
22 C1 1 PHO T6 22 0.00
[ bonds ]
; bonds between beads
; The bonds are finetuned against a backmapped gromos;
1 9 1 0.273 30000.0
9 10 1 0.298 23000.0
1 10 1 0.318 25000.0
1 10 1 0.318 25000.0
3 11 1 0.274 25000.0
3 12 1 0.319 23000.0
3 12 1 0.29 21000.0
11 12 1 0.29 21000.0
5 13 1 0.24 25000.0
5 13 1 0.24 25000.0
5 14 1 0.355 21000.0
5 14 1 0.355 21000.0
6 14 1 0.28 25000.0
6 14 1 0.28 25000.0
13 14 1 0.29 22000.0
13 14 1 0.29 22000.0
```

Binding cofactors into your protein? → One ITP needed

```
; Name      Exclusions
PSII_Protein 1

[ atoms ]
;CHAIN A
  1 Qd      1 MET   BB   1      1 ; C
  2 C5      1 MET   SC1  2      0 ; C
  3 P5      2 THR   BB   3      0 ; C
  4 P1      2 THR   SC1  4      0 ; C
  5 P5      3 THR   BB   5      0 ; C
  6 P1      3 THR   SC1  6      0 ; C
  7 P5      4 THR   BB   7      0 ; C
  8 P1      4 THR   SC1  8      0 ; C
  9 P5      5 LEU   BB   9      0 ; S
 10 C1      5 LEU   SC1 10      0 ; S
 11 P5      6 GLN   BB   11     0 ; C
PSII.itp
 770 P5      343 LEU   BB   770     0 ; C
 771 C1      343 LEU   SC1  771     0 ; C
 772 Qa      344 ALA   BB   772     -1 ; C
;CHAIN B
 773 Qd      2 GLY   BB   773     1 ; C
 774 P5      3 LEU   BB   774     0 ; C
 775 C1      3 LEU   SC1  775     0 ; C
 776 P4      4 PRO   BB   776     0 ; C
 777 C3      4 PRO   SC1  777     0 ; C
 778 P5      5 TRP   BB   778     0 ; C
 779 SC4     5 TRP   SC1  779     0 ; C
 780 SNd     5 TRP   SC2  780     0 ; C
 781 SC5     5 TRP   SC3  781     0 ; C
 782 SC5     5 TRP   SC4  782     0 ; C
 783 P5      6 TYR   BB   783     0 ; C
 784 SC4     6 TYR   SC1  784     0 ; C
 785 SC4     6 TYR   SC2  785     0 ; C
PSII.itp
5775 P5      38 GLN   BB   5775     0 ; C
5776 P4      38 GLN   SC1  5776     0 ; C
5777 P5      39 ARG   BB   5777     0 ; C
5778 N0      39 ARG   SC1  5778     0 ; C
5779 Qd      39 ARG   SC2  5779     1 ; C
5780 Qa      40 SER   BB   5780     -1 ; C
5781 P1      40 SER   SC1  5781     0 ; C
;CHAIN Z
5782 Qd      1 MET   BB   5782     1 ; C
5783 C5      1 MET   SC1  5783     0 ; C
5784 Nd      2 THR   BB   5784     0 ; 1
5785 P1      2 THR   SC1  5785     0 ; 1
5786 Nd      3 ILE   BB   5786     0 ; 1
5787 C1      3 ILE   SC1  5787     0 ; 1
5788 Nd      4 LEU   BB   5788     0 ; 1
5789 C1      4 LEU   SC1  5789     0 ; 1
PSII.itp
/CHA
```

```
5892 C1      52 LEU   SC1  5892     0 ; H
5893 N0      53 VAL   BB   5893     0 ; H
5894 C2      53 VAL   SC1  5894     0 ; H
5895 Na      54 VAL   BB   5895     0 ; 2
5896 C2      54 VAL   SC1  5896     0 ; 2
5897 Na      55 GLY   BB   5897     0 ; 2
5898 Na      56 VAL   BB   5898     0 ; 2
5899 C2      56 VAL   SC1  5899     0 ; 2
5900 Na      57 LEU   BB   5900     0 ; 2
5901 C1      57 LEU   SC1  5901     0 ; 2
5902 P5      58 ASN   BB   5902     0 ; C
5903 P5      58 ASN   SC1  5903     0 ; C
5904 P5      59 PHE   BB   5904     0 ; C
5905 SC5     59 PHE   SC1  5905     0 ; C
5906 SC5     59 PHE   SC2  5906     0 ; C
5907 SC5     59 PHE   SC3  5907     0 ; C
5908 P5      60 PHE   BB   5908     0 ; C
5909 SC5     60 PHE   SC1  5909     0 ; C
5910 SC5     60 PHE   SC2  5910     0 ; C
5911 SC5     60 PHE   SC3  5911     0 ; C
5912 Nda     61 VAL   BB   5912     0 ; T
5913 C2      61 VAL   SC1  5913     0 ; T
5914 Qa     62 VAL   BB   5914     -1 ; C
5915 C2      62 VAL   SC1  5915     0 ; C
=====
;CLA
5916 SP3     604 CLA   NA   5916     -0.25
5917 SC3     604 CLA   CHA  5917     0
5918 SP3     604 CLA   NB   5918     -0.25
5919 SC3     604 CLA   CHB  5919     0
5920 SP3     604 CLA   NC   5920     -0.25
5921 SC3     604 CLA   CHC  5921     0
5922 SP3     604 CLA   ND   5922     -0.25
5923 SC3     604 CLA   CHD  5923     0
5924 SC3     604 CLA   C1A  5924     0
5925 SC3     604 CLA   C2A  5925     0
5926 SC3     604 CLA   C1B  5926     0
5927 SC3     604 CLA   C2B  5927     0
5928 SC4     604 CLA   C1C  5928     0
5929 SNa     604 CLA   C2C  5929     0
5930 Na      604 CLA   T1   5930     0
5931 SC3     604 CLA   C1D  5931     0
5932 SC3     604 CLA   C2D  5932     0
5933 Na      604 CLA   T2   5933     0
5934 C3      604 CLA   T3   5934     0
5935 C1      604 CLA   T4   5935     0
5936 C1      604 CLA   T5   5936     0
5937 C1      604 CLA   T6   5937     0
5938 S00     604 CLA   MG   5938     1
5939 SP3     607 CLA   NA   5939     -0.25
5940 SC3     607 CLA   CHA  5940     0
5941 SP3     607 CLA   NB   5941     -0.25
```

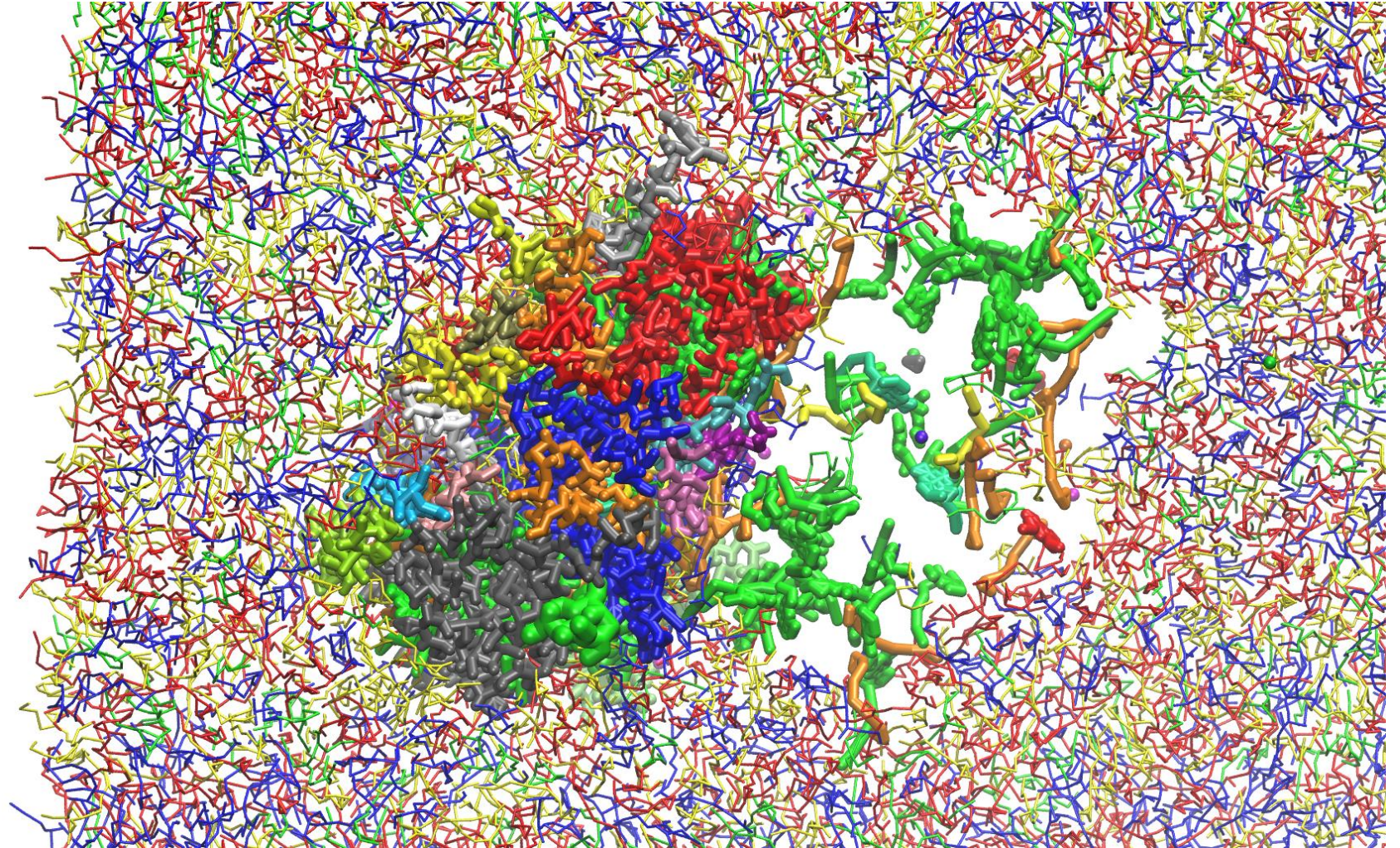
```
6750 SC3     642 HEM   CHC   6750     0
6751 SP1     642 HEM   ND   6751     -0.1
6752 SC3     642 HEM   CHD   6752     0
6753 SC3     642 HEM   C2A   6753     0
6754 SC3     642 HEM   C3A   6754     0
6755 SC3     642 HEM   C2B   6755     0
6756 SC3     642 HEM   C3B   6756     0
6757 SC3     642 HEM   C1C   6757     0
6758 SC3     642 HEM   C2C   6758     0
6759 Qa      642 HEM   C3C   6759     -1
6760 SC3     642 HEM   C1D   6760     0
6761 SC3     642 HEM   C2D   6761     0
6762 Qa      642 HEM   C3D   6762     -1
6763 SQ0     642 HEM   FE   6763     0.4
;OEX
6764 Q0      601 OEX   XCA   6764     1.5 ; the tot
oxyl groups, of which the charges are neutralized in vivo by the O
6765 Q0      601 OEX   X1   6765     1.5
6766 Q0      601 OEX   X2   6766     1.5
6767 Q0      601 OEX   X3   6767     1.5
;FE
6768 Q0      603 FE2   FE   6768     +2
;BCT
6769 Qda     681 BCT   BCT   6769     -1
;CA
6770 Q0      767 CA   CA   6770     +2
6771 Q0      796 CA   CA   6771     +2
6772 Q0      803 CA   CA   6772     +2
;MG
6773 Q0      771 MG   MG   6773     +2
;CL
6774 Q0      679 CL   CL-  6774     -1
6775 Q0      680 CL   CL-  6775     -1
6776 Q0      808 CL   CL-  6776     -1
;PHEOPHYTINS
6777 SP3     608 PHO   NA   6777     0
6778 SC3     608 PHO   CHA  6778     0
6779 SP3     608 PHO   NB   6779     0
6780 SC3     608 PHO   CHB  6780     0
6781 SP3     608 PHO   NC   6781     0
6782 SC3     608 PHO   CHC  6782     0
6783 SP3     608 PHO   ND   6783     0
6784 SC3     608 PHO   CHD  6784     0
6785 SC3     608 PHO   C1A  6785     0
6786 SC3     608 PHO   C2A  6786     0
6787 SC3     608 PHO   C1B  6787     0
6788 SC3     608 PHO   C2B  6788     0
6789 SC4     608 PHO   C1C  6789     0
6790 SNa     608 PHO   C2C  6790     0
6791 Na      608 PHO   T1   6791     0
6792 SC3     608 PHO   C1D  6792     0
6793 SC3     608 PHO   C2D  6793     0
```

Everything ready?

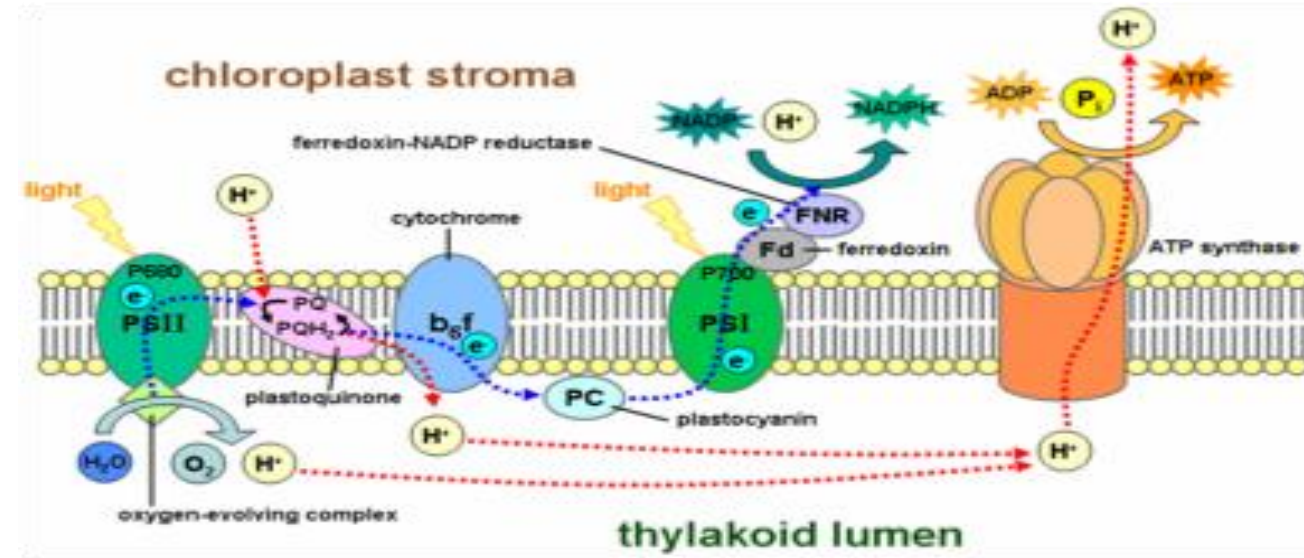
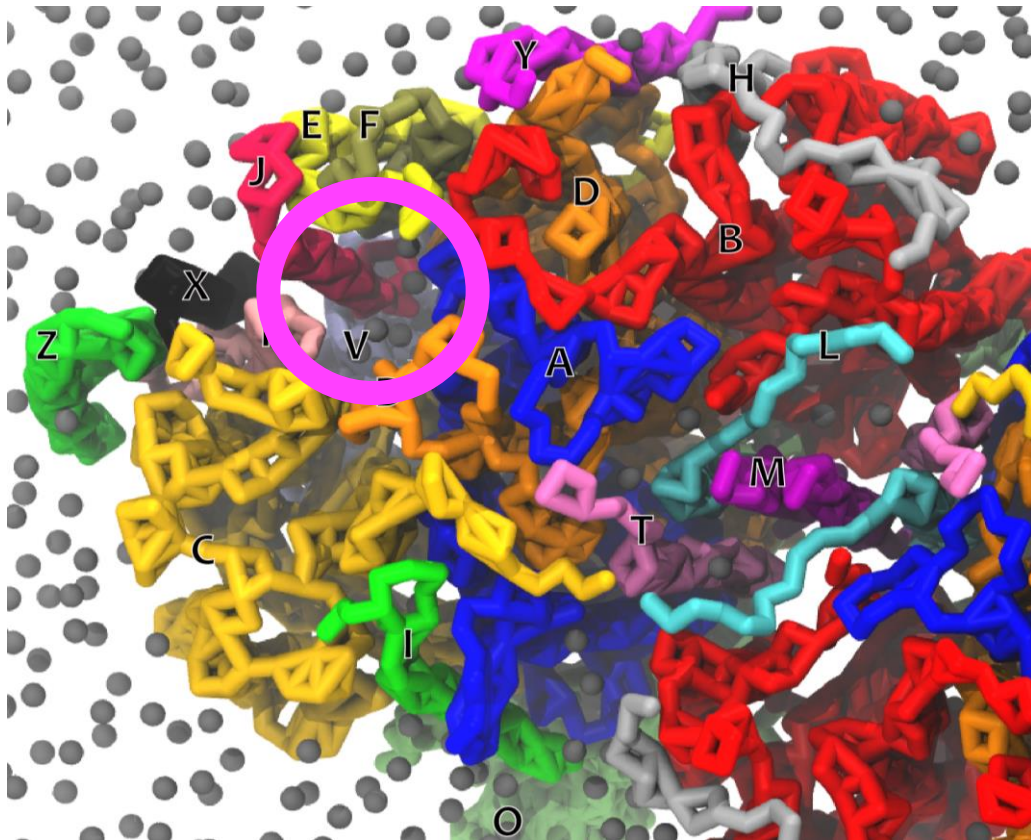
- Good energy minimization
 - Multiple rounds
- Careful equilibration
 - Slowly increasing
 - Time step
 - temperature

Where should i look at?

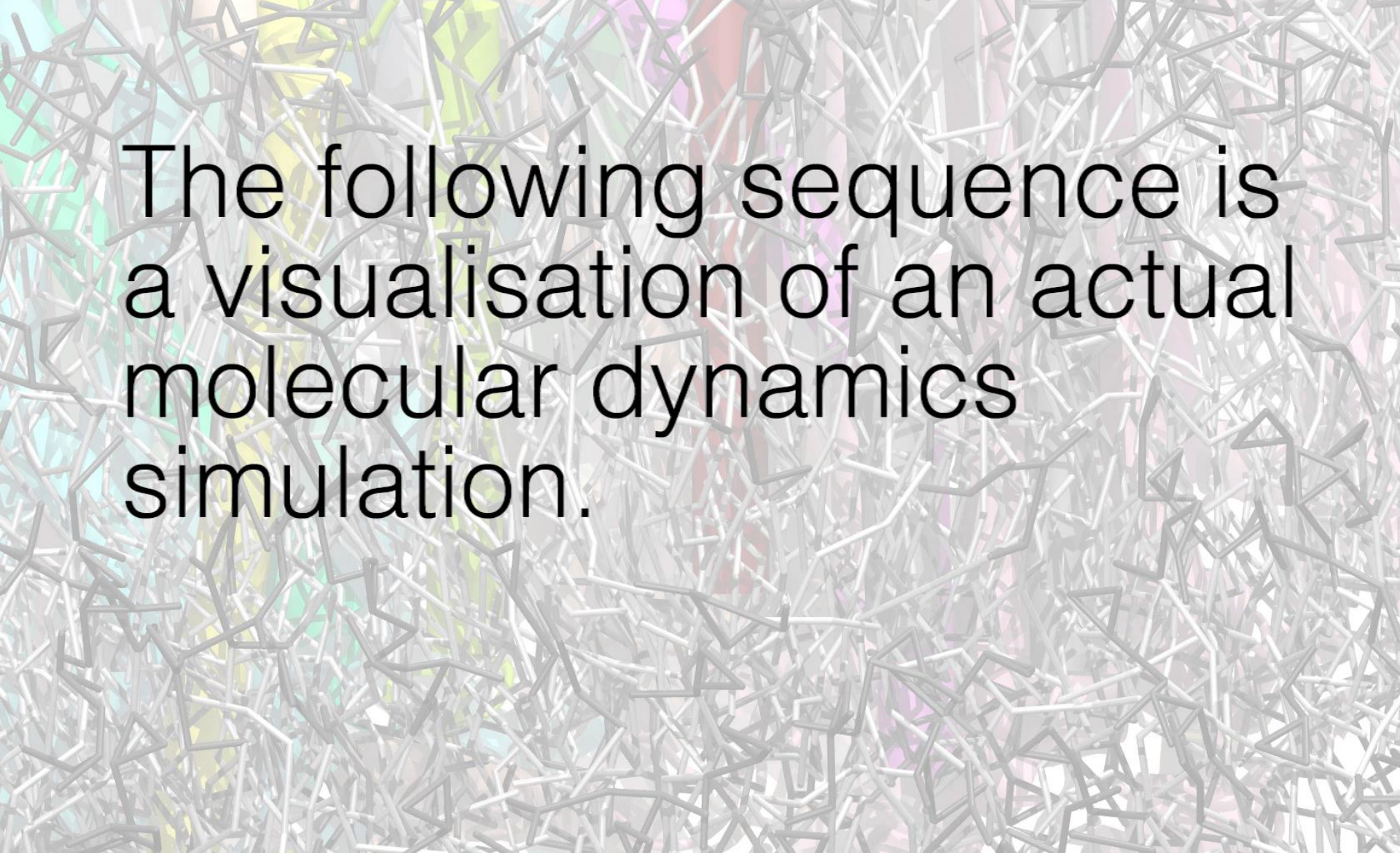
- [MOVIE PSII TOP]
- Nothing seems to happen
- /to much to look at.



PLQ exchange cavity



Analysis



The following sequence is a visualisation of an actual molecular dynamics simulation.

Counting PLQs entering/leaving PSII

- PLQs are long molecules
- Entry/exit is slow
- PLQs can enter partly and then go back
- PLQ can be in 2 channels at the same time
- **Channels move over time**

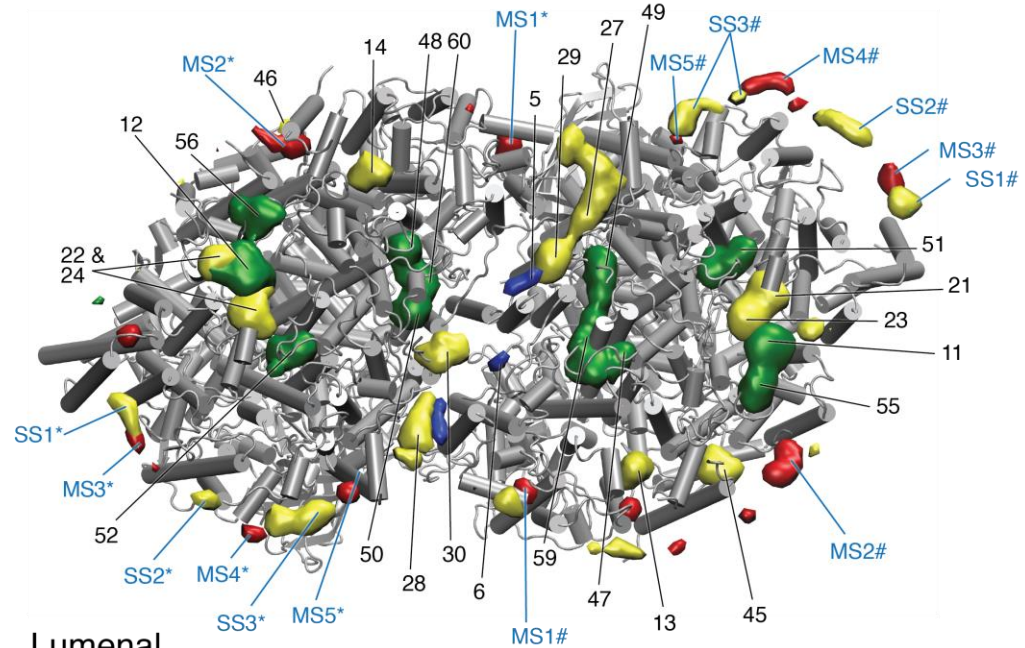
Track the PLQs on a bead level

- For every PLQ bead, each frame:
 - Is it in contact with any of the three channels
 - If it is inside or outside the cavity
- For every PLQ
 - If more than 70% of its beads moved through the same channel
 - entry/exit event

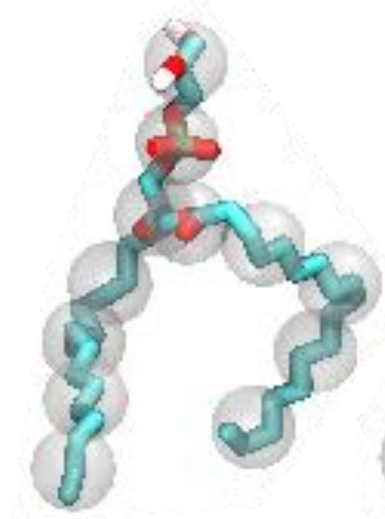
PLQ ENTERING MOVIE

Lipid binding sites

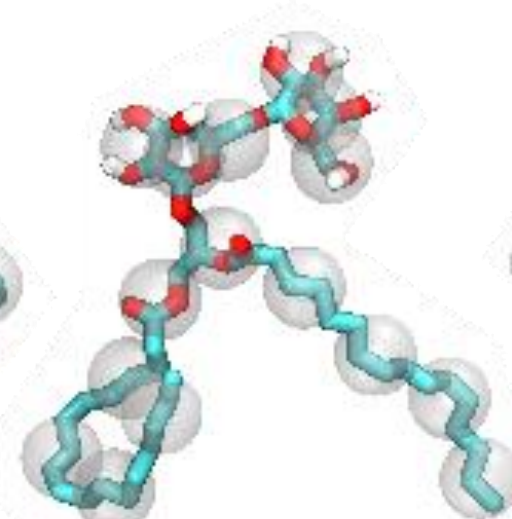
Stromal



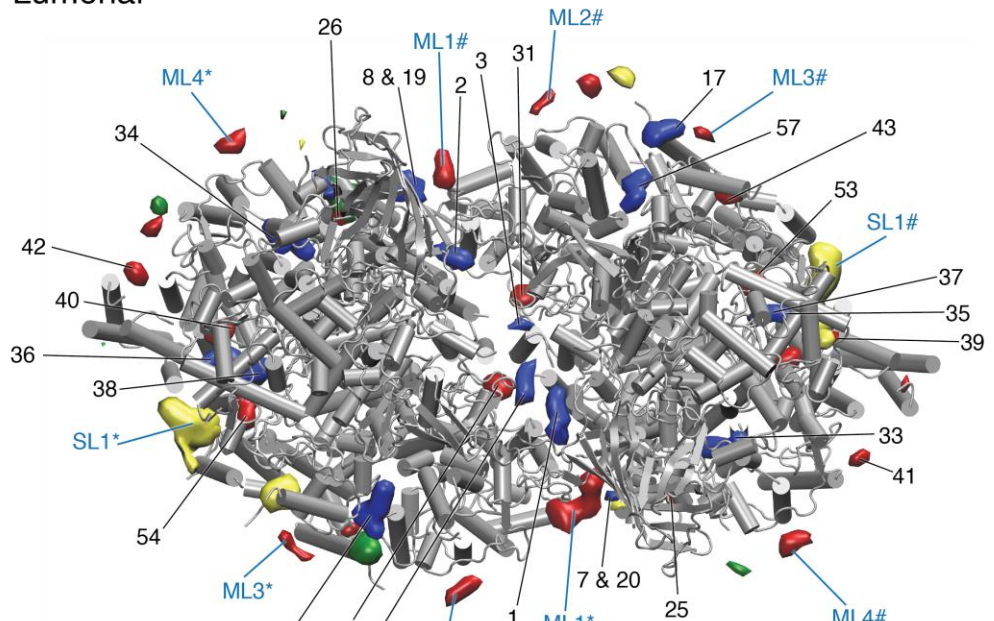
PG



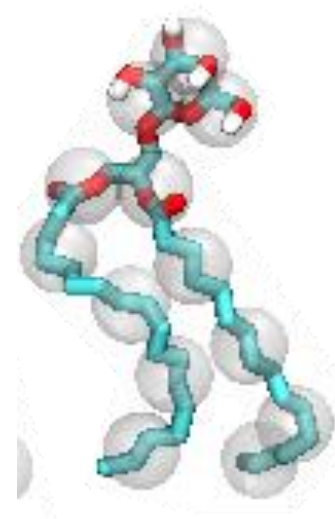
DGDG



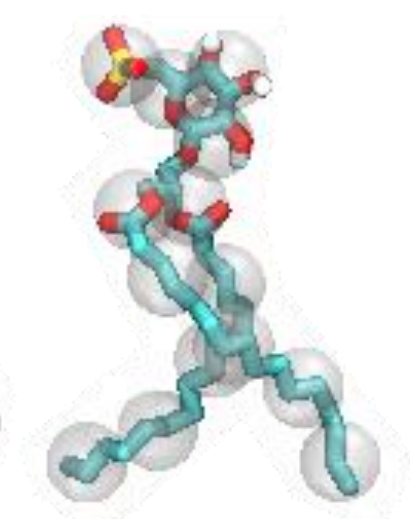
Luminal



MGDG

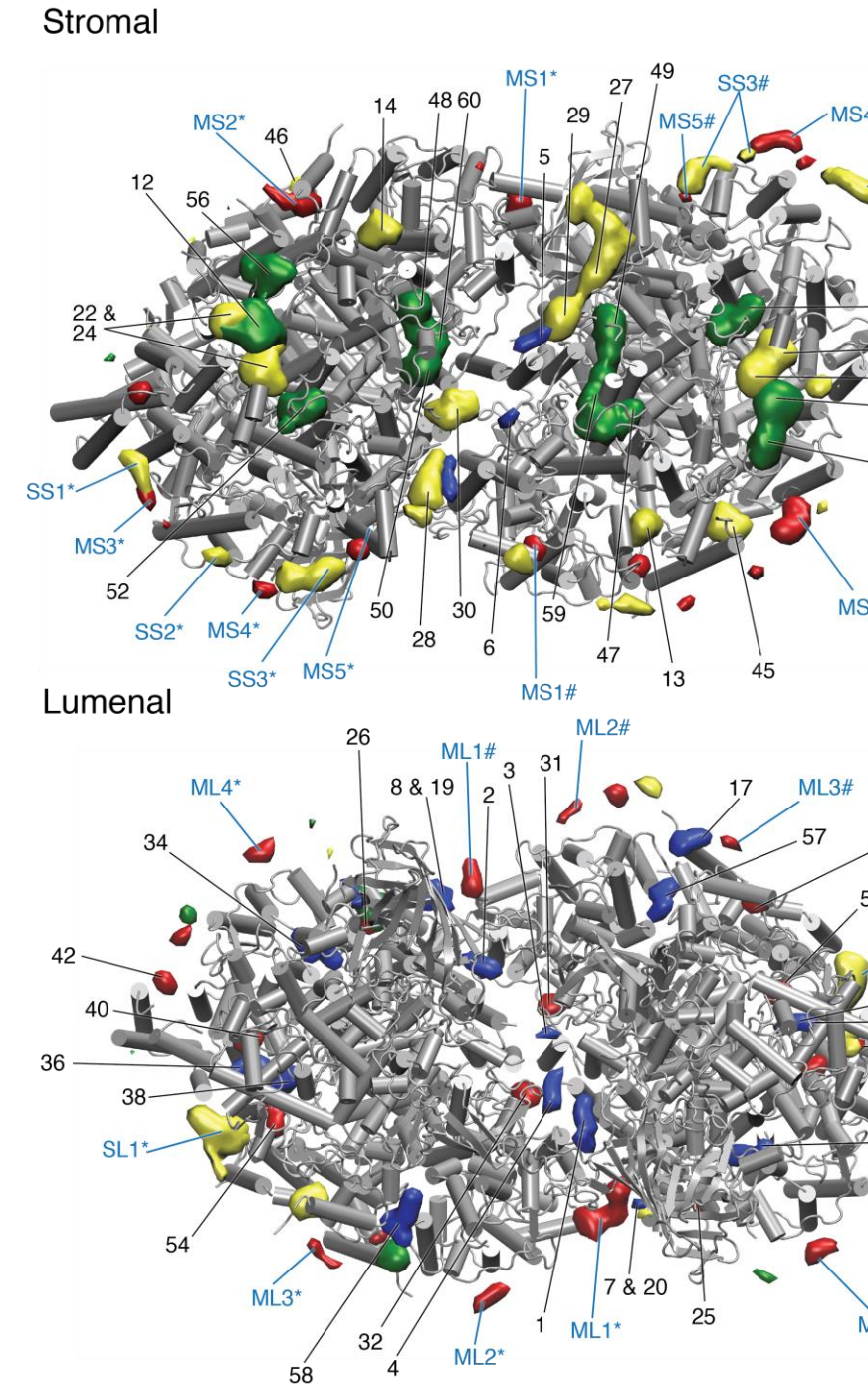


SQDG



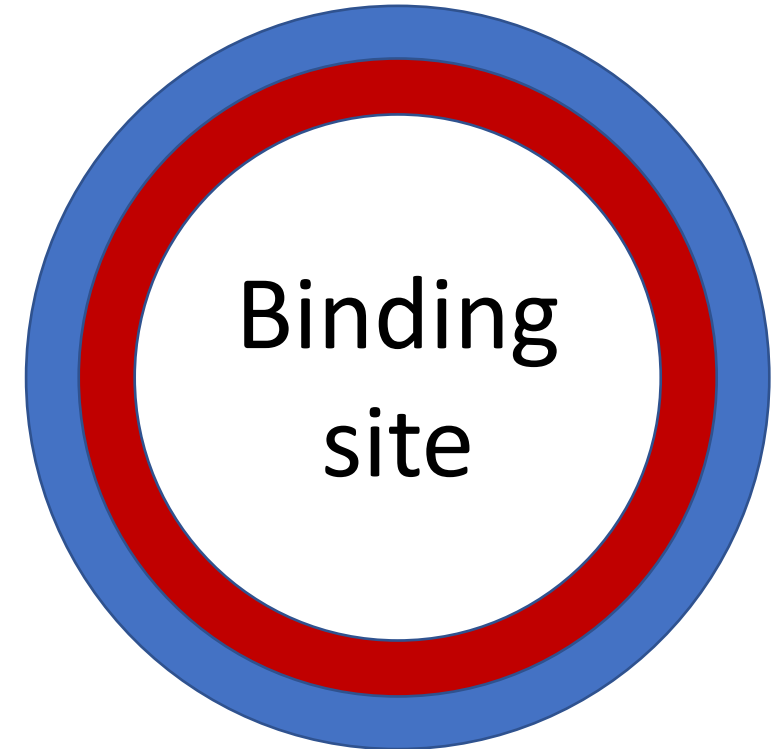
How to define a binding site

- Define 'phrases'
 - Amino acids a lipid is in contact with at a frame
 - Total number of phrases = number of lipids x frames
- Cluster phrases
- From clusters define binding sites



Residence times

- Double cut off method → removes noise
- Lipid is in contact with binding site
 - Once it enters red ring
- Lipid is not in contact anymore
 - When leaving blue ring



Conclusion

- Carefully prepare your system, it will take a long time!
- Look a lot at your simulations:
 - Only protein
 - Protein with lipids
 - Protein cofactors
 - Cofactors lipids
- Team up with smart colleagues (e.g. writing scripts)
- Make descisions
 - Often there is not a right way to do things